

## Formation of Fusaric Acid by Fungi of the Genus *Fusarium*

W.-U. Mutert, H. Lütfring, and W. Barz

Lehrstuhl für Biochemie der Pflanzen der Universität Münster, Hindenburgplatz 55, D-4400 Münster

D. Strack

Botanisches Institut der Universität zu Köln

Z. Naturforsch. **36 c**, 338–339 (1981);  
received January 19, 1981

Fusaric Acid, *Fusarium*, *Gibberella*, Biosynthesis, High Performance Liquid Chromatography

Among various *Fusarium* strains tested *Gibberella fujikuroi* (SAW) WR was shown to be a high producer of the phytotoxin fusaric acid.

During studies on phytopathogenic *Fusarium* fungi [1, 2] and their effects on plant cell cultures [3, 4] it became necessary to determine in our experimental strains [1] the capacity for fusaric acid (5-*n*-butyl-pyridine-2-carboxylic acid) formation. This phytotoxin has repeatedly been isolated from various *Fusarium* species of both the *Elegans* and the *Martiella* group as well as of *Gibberella fujikuroi* [5, 6]. The involvement of fusaric acid in wilt diseases of various plant species has conclusively been shown [7]. Action mechanism and metabolism of fusaric acid in plant tissues warrants further investigations which have been started by using cell cultures of host plants [3, 4] and fusaric acid producing fungal cultures.

Strains were grown in shake culture [1] and at intervals between 1 and 28 days of fungal growth, aliquots of the culture fluid of two vials each were extracted with ethylacetate at pH 3.8 [6]. The phytotoxin was quantitatively determined in these aliquots by high performance liquid chromatography. This very accurate procedure (LiChrosorb RP-8 column) well separated fusaric acid from other UV absorbing metabolites and in contrast to other chromatographic procedures [3] allowed numerous analyses in a short period of time. In addition, fusaric acid was isolated, crystallized from *n*-hexane and identified by UV ( $\lambda_{\max}$  226 and 269 nm),  $^1\text{H-NMR}$  (signals at 12.16 (s), 8.75 (s), 8.25 (d) and 7.82 (d), 2.84 (t), 1.58 (m) and 0.99 (t) ppm) and MS ( $m/e$   $M^+$  179, 162, 135 (100), 119, 106, 91, 77, 65 and 27) spectroscopy. Evi-

dence for the occurrence of dehydrofusaric acid has not been obtained in the shake cultures.

Strains III (*F. oxysporum* Schlecht ex Fr. f. *medicaginis*, CBS 179.29), XIV (*Gibberella sanbinetti*, CBS 265.54) and XVIII (*F. oxysporum* f. sp. *pisi*, recently isolated from infected "Picadir" pea plants) were shown (TLC,  $S_2$  and  $S_3$ ) not to synthesize fusaric acid. Strains X (*F. oxysporum* Schlecht ex Fr. f. *apii*, CBS 184.38) and XVII (*F. oxysporum* f. sp. *pisi*, also isolated from infected "Picadir" pea plants) produced moderate amounts of fusaric acid between 290 and 12 mg/l culture fluid (by HPLC). Strain XIII (*Gibberella fujikuroi* (Saw) WR, CBS 186.56) turned out to be one of the best producers of fusaric acid reported so far [8] because up to 2550 mg/l culture medium were measured. With the exception [8] of *G. fujikuroi* (Saw) Woll, strain ETH M 82, the yield of fusaric acid in most previous studies ranged between 100–1000 mg/l. As shown in Fig. 1 fusaric acid formation occurred parallel to growth with maximum production between days 3 and 8. In contrast to other reports [9] further metabolism of fusaric acid upon longer incubation periods could not be observed in our studies.

Attempts to induce fusaric acid formation in strain XVIII or improve its accumulation in strains X, XIII and XVII by growth at increased zinc levels (0.02–10 ppm) failed though earlier reports [10] had described the stimulatory role of zinc ions.

Our strain of *Gibberella fujikuroi* was also used to synthesize larger quantities of  $^{14}\text{C}$ -labelled fusaric acid using [ $^{14}\text{C}$ ]acetate [11, 12]. With reference to Fig. 1 and based on preliminary studies [4] using varied precursor concentrations, 8 flask (total volume 800 ml) were incubated with 800  $\mu\text{Ci}$  [ $^{14}\text{C}$ ]acetate ( $10^{-3}$  M) for 72 h between day 3 and day 6 of the growth curve. Radiochemically pure fusaric acid was recovered by ethylacetate extraction, repeated crystallisation from *n*-hexane and prep. TLC (yield: 1.064 g; spec. radioact.: 13.3  $\mu\text{Ci}/\text{mmol}$ ; total precursor incorporation: 10%). In general, the data show the great diversity for phytotoxin formation among *Fusarium* fungi [6, 7] and its great variation in total yield.

## Experimental

### Fungi

Cultivation and growth (100 ml batches) of fungal strains were as previously described [1].

Reprint requests to Prof. Dr. W. Barz.

0341-0382/81/0300-0338 \$ 01.00/0



Dieses Werk wurde im Jahr 2013 vom Verlag Zeitschrift für Naturforschung in Zusammenarbeit mit der Max-Planck-Gesellschaft zur Förderung der Wissenschaften e.V. digitalisiert und unter folgender Lizenz veröffentlicht: Creative Commons Namensnennung-Keine Bearbeitung 3.0 Deutschland Lizenz.

Zum 01.01.2015 ist eine Anpassung der Lizenzbedingungen (Entfall der Creative Commons Lizenzbedingung „Keine Bearbeitung“) beabsichtigt, um eine Nachnutzung auch im Rahmen zukünftiger wissenschaftlicher Nutzungsformen zu ermöglichen.

This work has been digitalized and published in 2013 by Verlag Zeitschrift für Naturforschung in cooperation with the Max Planck Society for the Advancement of Science under a Creative Commons Attribution-NoDerivs 3.0 Germany License.

On 01.01.2015 it is planned to change the License Conditions (the removal of the Creative Commons License condition "no derivative works"). This is to allow reuse in the area of future scientific usage.

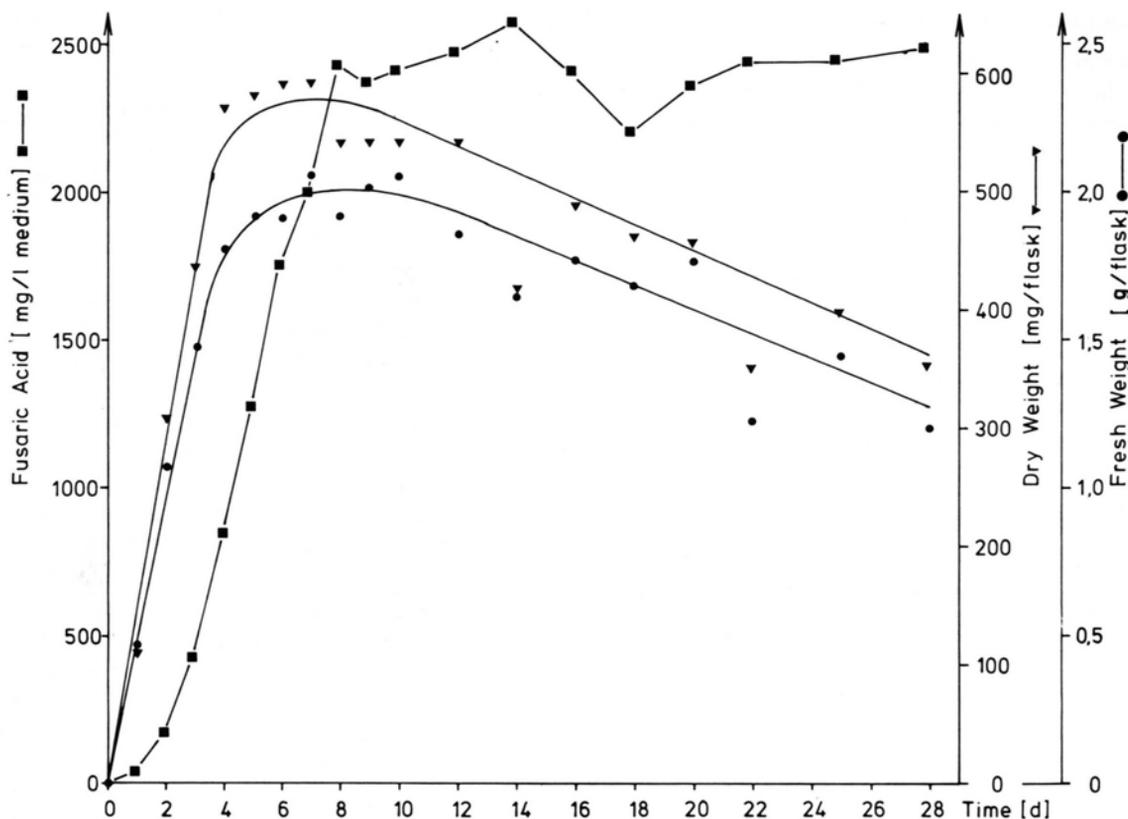


Fig. 1. Growth of *Gibberella fujikuroi* (strain XIII, CBS 186.56) in shake culture and accumulation of fusaric acid.

### Chromatography

TLC on silica gel plates was performed with solvent systems  $S_1$ :  $\text{CH}_3\text{OH}:\text{CHCl}_3:\text{H}_2\text{O}$  3:2:1;  $S_2$ : iso-PrOH: $\text{H}_2\text{O}$  85:15 and  $S_3$ : *n*-But-OH:HOAc: $\text{H}_2\text{O}$  4:1:1. Fusaric acid was detected with Dragendorff-reagent or UV absorption.

### HPLC

Spectra-Physics chromatograph with a Spectro Flow SF 770 UV detector (270 nm), a LiChrosorb RP-8 (5  $\mu\text{m}$ ) column (250  $\times$  4 mm). Solvents A: 1.5%  $\text{H}_3\text{PO}_4$  in  $\text{H}_2\text{O}$ ; B: 1.5%  $\text{H}_3\text{PO}_4$  + 20%  $\text{CH}_3\text{CO}_2\text{H}$  + 25%  $\text{CH}_3\text{CN}$  in  $\text{H}_2\text{O}$ ; column development isocratic with 50% B in A. Flow 1 ml/min and injection volume 20  $\mu\text{l}$ . Alternatively, a LiChrosorb-NH<sub>2</sub> (5  $\mu\text{m}$ ) column (125  $\times$  4 mm) with solvent 0.5%  $\text{H}_3\text{PO}_4$  + 2.5%  $\text{H}_2\text{O}$  in  $\text{CH}_3\text{CN}$  was used. The analytical procedure was quantitated with authentic fusaric acid (Sigma).

### Acknowledgements

Financial support by Minister für Wissenschaft und Forschung des Landes Nordrhein-Westfalen and Deutsche Forschungsgemeinschaft is gratefully acknowledged.

- [1] W. Barz, R. Schlepphorst, and J. Laimer, *Phytochemistry* **15**, 87 (1976).
- [2] K.-M. Weltring and W. Barz, *Z. Naturforsch.* **35 c**, 399 (1980).
- [3] W.-U. Mutert, Diplomarbeit Universität Münster 1980.
- [4] H. Lütfring, Staatsexamensarbeit Universität Münster 1981.
- [5] K. Rudolph, *Physiological Plant Pathology* (R. Heitefuß and P. H. Williams, eds.) pp. 296 ff., Springer-Verlag Berlin, Heidelberg, New York 1976.
- [6] N. Claydon, J. F. Grove, and M. Pople, *Phytochemistry* **16**, 603 (1977).
- [7] H. Kern, *Phytotoxins in Plant Diseases* (R. K. S. Wood, H. Ballio, and A. Graniti, eds.) p. 39 ff., Academic Press, London and New York 1972.
- [8] R. S. Sandhu, *Phytopath. Z.* **37**, 33 (1959).
- [9] R. Braun, *Phytopath. Z.* **39**, 197 (1960).
- [10] T. A. Egli, *Phytopath. Z.* **66**, 223 (1969).
- [11] R. D. Hill, A. M. Unrau, and D. T. Canvin, *Canad. J. Chem.* **44**, 2077 (1966).
- [12] D. Desaty, A. G. McInnes, D. G. Smith, and L. C. Vining, *Can. J. Biochem.* **46**, 1293 (1968).